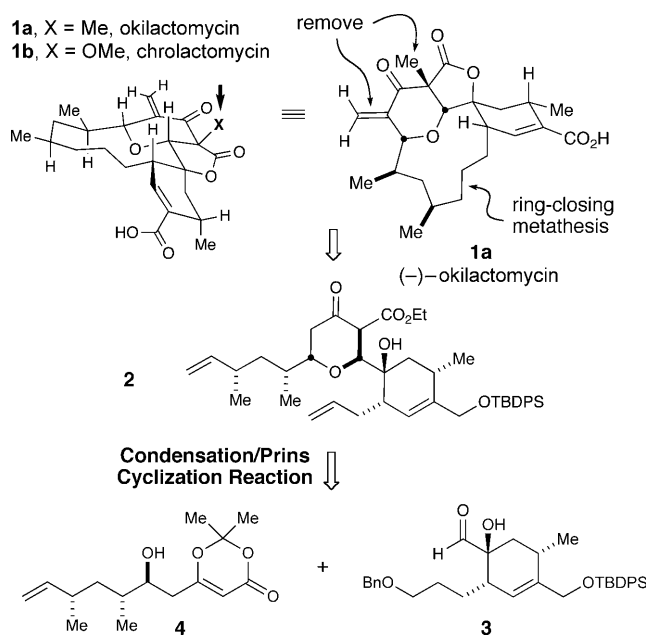


Synthesis of (–)-Okilactomycin by a Prins-Type Fragment-Assembly Strategy**

Jason M. Tenenbaum, William J. Morris, Daniel W. Custar, and Karl A. Scheidt*

Okilactomycin (**1a**) is a structurally interesting antitumor antibiotic that was isolated from *Streptomyces griseoflavus* in 1987.^[1] In vitro studies have demonstrated that **1a** exhibits significant antitumor and antiproliferative activity against both lymphoid leukemia L1210 cells and P388 leukemia cells with IC₅₀ values of 216 nM and 89 nM, respectively.^[1b] A closely related compound, chrolactomycin (**1b**), differs only in structure by the replacement of a methyl group with a methoxy moiety at the pyranone/lactone ring fusion and displays promising telomerase inhibition.^[1c,d] In addition to their potent biological activity, these compounds possess a compact and intriguing architecture. The tricyclic core is characterized by a unique 6,5-fused tetrahydropyranone γ -lactone bicycle with a spiro fusion to a highly substituted cyclohexene. A strained deoxygenated dipropionate segment spans this unusual tricycle to generate a highly rigid tetracyclic topology. Despite the biological activity and structural complexity, there have been only limited reports on the synthesis of okilactomycin (**1a**) over the last two decades, namely from the laboratories of Takeda, Paquette, and Smith.^[2] These synthetic efforts culminated in a total synthesis of unnatural enantiomer (–)-**1a** and determination of the absolute configuration of the natural product by Smith et al. in 2007.^[2d,e] There are no syntheses of chrolactomycin (**1b**) reported to date. We disclose herein a convergent synthesis of (–)-**1a** utilizing a Prins-type Maitland–Japp cyclization strategy of two advanced fragments.

Our retrosynthetic plan is outlined in Scheme 1. Given the electrophilic nature of the exomethylene unit, we elected for a late-stage installation of this moiety. We envisioned that the key tetracyclic precursor could be accessed from the lactonization of seco-ester **2**, and subsequent ring-closing metathesis



Scheme 1. Retrosynthetic strategy. TBDPS = *tert*-butyldiphenylsilyl.

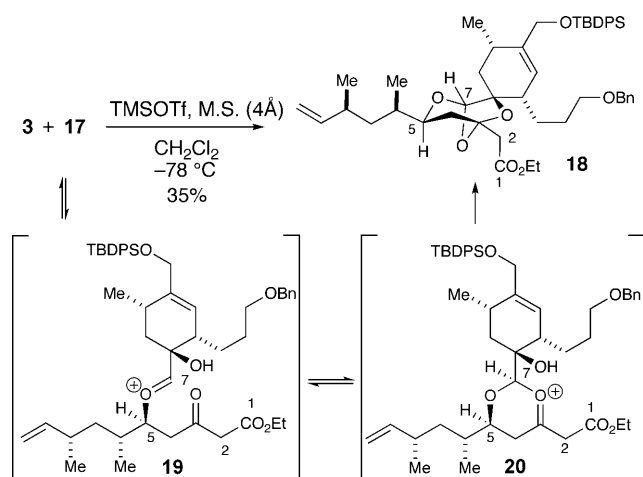
(RCM) to install the 11-membered macrocycle. The tetrahydropyranone heart of the molecule would be formed from a convergent union of the corresponding α -hydroxy aldehyde **3** and β -hydroxy dioxinone **4** through a Prins cyclization.^[3] The cyclohexenyl aldehyde could be accessed using an asymmetric Diels–Alder reaction in conjunction with functional group manipulation and Rubottom oxidation. The β -hydroxy dioxinone motif could be constructed using a vinylogous aldol reaction of an acetoacetate equivalent.

The synthesis of aldehyde **3** centered on an *endo*-selective Diels–Alder reaction to install the necessary substitution pattern. The requisite diene for this [4+2] strategy was constructed starting with the hydrozirconation/iodination reaction of benzyl-protected alkyne **5** (Scheme 2).^[4] A lithium–halogen exchange of vinyl iodide **6** with *n*BuLi and subsequent treatment of the resulting vinyllithium species with Weinreb amide **7** afforded the desired enone in 65% yield as a >20:1 mixture of *E/Z* isomers. A selective Wittig olefination with ethyltriphenylphosphonium bromide provided diene **8** with >20:1 *E/Z* selectivity.^[5,6] In the first key step of the synthesis, the core cyclohexene was formed in 86% yield with 20:1 diastereoselectivity for the *endo* product through the Diels–Alder reaction of diene **8** (1 equiv) with acrylimide **9** (1.1 equiv).^[7,8] Early attempts using dialkyl aluminum halides or alternative Lewis acids for this cycloaddition with realistic levels of diene (i.e. <10 equiv) resulted

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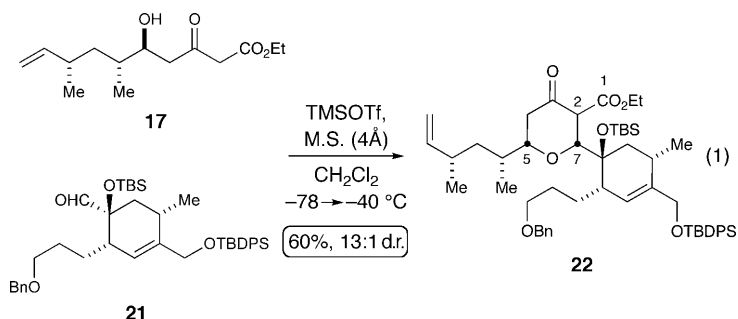
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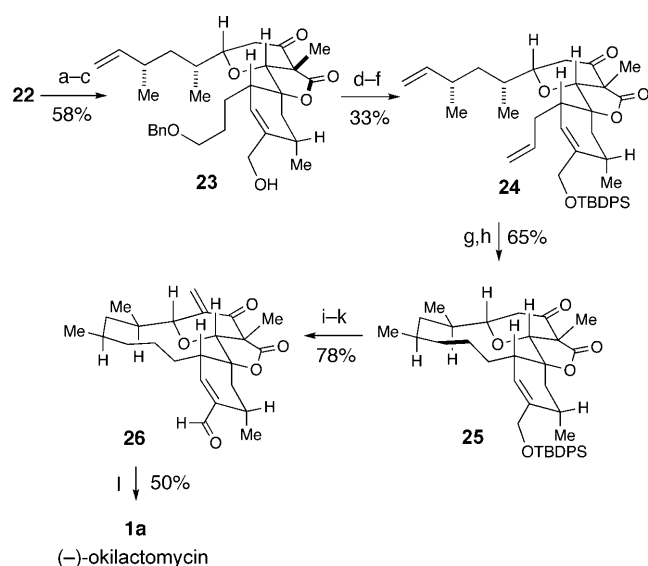
Scheme 4. Proposed pathway for the formation of **18**. M.S. = molecular sieves.

Despite the unexpected formation of tricycle **18**, we were encouraged that δ -hydroxy- β -ketoester **17** was a competent coupling partner for the sterically demanding α -hydroxy aldehyde **3**. A revised strategy was pursued involving the protection of the tertiary alcohol in **3** to facilitate the desired cyclization pathway. After extensive experimentation with various protecting groups, only TBS ether **21** survived the Lewis acid mediated conditions. Ultimately, the optimal conditions for the stereoselective coupling of **21** and **17** were TMSOTf in CH_2Cl_2 , and led to the desired tetrahydropyranone **22** in a 60% yield and as a 13:1 mixture of diastereomers favoring the desired 2,6-*cis* isomer [Eq. (1)].



The union of these two fragments establishes the key C–O and C–C bonds of the central 6,5-fused pyranone/lactone in a single operation. In addition, this process relays the configuration of β -ketoester **17** to the new stereogenic center at C7 with high level of fidelity and constructs the full carbon skeleton of the natural product.

The global removal of the silyl groups of pyranone **22** was accomplished with aqueous HF and set the stage for completion of the synthesis. A lactonization between the unmasked hindered tertiary alcohol and pendant ethyl ester with KOtBu and subsequent stereoselective C methylation produced the key tricycle **23** in a 58% yield with only one purification over the three-step sequence (Scheme 5). Allylic



Scheme 5. Completion of the synthesis of (–)-okilactomycin. Reagents and conditions: a) aq HF, CH_3CN . b) KOtBu, CH_2Cl_2 . c) K_2CO_3 , MeI, CH_3CN , 70 °C, 58% over three steps). d) TBDPSCl, imidazole, CH_2Cl_2 , 90%. e) DDQ (30 equiv), CH_2Cl_2 , 61%. f) 1. *o*-nitrophenyl-selenocyanate, PbU_3 , THF; 2. H_2O_2 , THF, 60%. g) Grubbs second-generation catalyst (40 mol %), CH_2Cl_2 , 40 °C. h) H_2 , PtO_2 , EtOAc, 65% over two steps. i) HF-py, THF, > 99% j) LiHMDS, dimethylmethylideneammonium iodide, THF 95%. k) Dess–Martin periodinane, CH_2Cl_2 , 83% l) NaClO_2 , NaH_2PO_4 , 2-methyl-2-butene, $t\text{BuOH}/\text{THF}/\text{H}_2\text{O}$ (4:4:1), 50%. DDQ = 2,3-dichloro-5,6-dicyanobenzoquinone.

alcohol **23** was reprotected as the TBDPS ether and the benzyl ether was cleaved under oxidative conditions with DDQ.^[17] The resultant primary alcohol was converted into the terminal olefin without interference from the additional double bond in the molecule by using the Grieco protocol.^[13]

At this stage, we pursued the planned ring-closing metathesis of **24** to form the macrocycle and complete the core of (–)-okilactomycin. This approach was originally proposed by Paquette^[2b,c] and subsequently realized in the Smith synthesis.^[2d] However, unique to our system was the presence of different functionality, most notably the olefin already installed in the cyclohexene ring. This internal alkene ultimately facilitates smooth installation of the α,β -unsaturated acid of the natural product (see below) and could have potentially compromised a metathesis approach.

Pleasingly, the exposure of bis(olefin) **24** to Grubbs second-generation catalyst and subsequent hydrogenation furnished the tetracycle **25** in 65% yield over the two steps.^[18] For our end-game approach, we were concerned that the exomethylene unit might be incompatible with conditions used to remove the silyl group, so we first removed the silyl unit and attempted to install the exocyclic olefin in the presence of the free allylic alcohol. Fortunately, silyl ether **25** was treated with HF-pyr to yield the allylic alcohol, and treatment with LiHMDS and dimethylmethylideneammonium iodide (Eschenmoser's salt) cleanly installed the exocyclic olefin.^[19] Finally, an oxidation to enal **26** with Dess–Martin periodinane (83% yield) followed by a Pinnick oxidation^[20] afforded (–)-

okilactomycin, which possessed identical characterization data (^1H NMR, ^{13}C NMR, HRMS, IR spectra) to the natural material.^[21] The $[\alpha]_{\text{D}}^{20}$ observed was $-20\text{ deg cm}^3\text{ g}^{-1}\text{ dm}^{-1}$ (MeOH, $c = 0.04\text{ g cm}^{-3}$) which is opposite in sign compared to isolated (+)-okilactomycin ($+34\text{ deg cm}^3\text{ g}^{-1}\text{ dm}^{-1}$, $c = 1.0\text{ g cm}^{-3}$, MeOH).^[2b,22]

In summary, the total synthesis of (–)-okilactomycin (**1a**) has been achieved in 1.0% overall yield over 26 steps as the longest linear sequence (via **17**). Stereoselective alkylation and Diels–Alder routes facilitated quick access to the δ -hydroxy β -ketoester and α -silyloxy aldehyde fragments, respectfully. A Lewis acid promoted Maitland–Japp reaction established the full carbon core with a high degree of diastereoselectivity for the 2,6-*cis* tetrahydropyrans core. This Prin-type transformation is one of the most advanced to date in terms of size and functionality of the reactants and further defines the potential of this approach for late-state unions of complex intermediates. The installation of the exocyclic olefin at the end of the synthesis and convergent nature of this route makes this synthesis amenable to the production of analogues and structure-activity relationship studies. The synthesis of these related compounds, which are intended for biological investigations, based on our complex fragment assembly Prins/Maitland–Japp route described here are ongoing in our laboratory.

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- [1] a) H. S. Imai, H. Kaniwa, T. Tokunaga, S. Fujita, T. Furuya, H. Matsumoto, M. Shimizu, *J. Antibiot.* **1987**, *40*, 1483–1489; b) H. S. Imai, K. I. Suzuki, M. Morioka, Y. Numasaki, S. Kadota, K. Nagai, T. Sato, M. Iwanami, T. Saito, *J. Antibiot.* **1987**, *40*, 1475–1482; c) R. Nakai, S. Kakita, A. Asai, S. Chiba, S. Akinaga, T. Mizukami, Y. Yamashita, *J. Antibiot.* **2001**, *54*, 836–838; d) R. Nakai, H. Ishida, A. Asai, H. Ogawa, Y. Yamamoto, H. Kawasaki, S. Akinaga, T. Mizukami, Y. Yamashita, *Chem. Biol.* **2006**, *13*, 183–189.
- [2] a) K. Takeda, A. Shimotani, E. Yoshii, K. Yamaguchi, *Heterocycles* **1992**, *34*, 2259–2261; b) S. L. Boulet, L. A. Paquette, *Synthesis* **2002**, 895–900; c) L. A. Paquette, S. L. Boulet, *Synthesis* **2002**, 888–894; d) A. B. Smith III, K. Basu, T. Bosanac, *J. Am. Chem. Soc.* **2007**, *129*, 14872–14874; e) A. B. Smith III, T. Bosanac, K. Basu, *J. Am. Chem. Soc.* **2009**, *131*, 2348–2358.
- [3] a) M. J. Cloninger, L. E. Overman, *J. Am. Chem. Soc.* **1999**, *121*, 1092–1093; b) W.-C. Zhang, G. S. Viswanathan, C.-J. Li, *Chem. Commun.* **1999**, 291–292; c) S. R. Crosby, J. R. Harding, C. D. King, G. D. Parker, C. L. Willis, *Org. Lett.* **2002**, *4*, 3407–3410; d) R. Jasti, J. Vitale, S. D. Rychnovsky, *J. Am. Chem. Soc.* **2004**, *126*, 9904–9905; e) K.-P. Chan, T.-P. Loh, *Org. Lett.* **2005**, *7*, 4491–4494; f) K.-P. Chan, Y. H. Ling, T.-P. Loh, *Chem. Commun.* **2007**, 939–941; g) F. Liu, T.-P. Loh, *Org. Lett.* **2007**, *9*, 2063–2066; h) I. M. Pastor, M. Yus, *Curr. Org. Chem.* **2007**, *11*, 925–957; i) L. J. Van Orden, B. D. Patterson, S. D. Rychnovsky, *J. Org. Chem.* **2007**, *72*, 5784–5793; j) K. B. Bahnck, S. D. Rychnovsky, *J. Am. Chem. Soc.* **2008**, *130*, 13177–13181; k) J. D. Elsworth, C. L. Willis, *Chem. Commun.* **2008**, 1587–1589.
- [4] a) D. W. Hart, J. Schwartz, *J. Am. Chem. Soc.* **1974**, *96*, 8115–8116; b) J. Schwartz, J. A. Labinger, *Angew. Chem.* **1976**, *88*, 402–409; *Angew. Chem. Int. Ed. Engl.* **1976**, *15*, 333–340.
- [5] B. E. Maryanoff, A. B. Reitz, *Chem. Rev.* **1989**, *89*, 863–927.
- [6] The *E/Z* ratio was determined by nOe interactions. See the Supporting Information for details.
- [7] a) D. A. Evans, K. T. Chapman, J. Bisaha, *J. Am. Chem. Soc.* **1984**, *106*, 4261–4263; b) D. A. Evans, K. T. Chapman, J. Bisaha, *J. Am. Chem. Soc.* **1988**, *110*, 1238–1256; c) G. Ho, D. Mathre, *J. Org. Chem.* **1995**, *60*, 2271–2273.
- [8] The absolute and relative configuration of **10** was determined by X-ray crystallographic analysis. CCDC 816131 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.
- [9] a) W. Roush, D. Barda, *J. Am. Chem. Soc.* **1997**, *119*, 7402–7403; b) W. Roush, D. Barda, C. Limberakis, R. Kunz, *Tetrahedron* **2002**, *58*, 6433–6454; c) W. R. Roush, C. Limberakis, R. K. Kunz, D. A. Barda, *Org. Lett.* **2002**, *4*, 1543–1546.
- [10] a) S. Hanessian, P. J. Murray, *Can. J. Chem.* **1986**, *64*, 2232–2234; b) L. K. Blasdel, A. G. Myers, *Org. Lett.* **2005**, *7*, 4281–4283.
- [11] J. Krüger, E. M. Carreira, *J. Am. Chem. Soc.* **1998**, *120*, 837–838.
- [12] W. Zhang, M. J. Robins, *Tetrahedron Lett.* **1992**, *33*, 1177–1180.
- [13] P. A. Grieco, S. Gilman, M. Nishizawa, *J. Org. Chem.* **1976**, *41*, 1485–1486.
- [14] W. J. Morris, D. W. Custar, K. A. Scheidt, *Org. Lett.* **2005**, *7*, 1113–1116.
- [15] a) P. A. Clarke, W. H. C. Martin, *Org. Lett.* **2002**, *4*, 4527–4529; b) P. A. Clarke, W. H. C. Martin, J. M. Hargreaves, C. Wilson, A. J. Blake, *Chem. Commun.* **2005**, 1061–1063; c) P. A. Clarke, W. H. C. Martin, J. M. Hargreaves, C. Wilson, A. J. Blake, *Org. Biomol. Chem.* **2005**, *3*, 3551–3563; d) F. R. Japp, W. Maitland, *J. Chem. Soc. Trans.* **1904**, 85, 1473–1489.
- [16] The configuration was assigned by an nOe interactions.
- [17] N. Ikemoto, S. L. Schreiber, *J. Am. Chem. Soc.* **1992**, *114*, 2524–2536.
- [18] For a related ring-closing metathesis strategy and execution, see references. [2d] and [2e].
- [19] a) J. Schreiber, H. Maag, N. Hashimoto, A. Eschenmoser, *Angew. Chem.* **1971**, *83*, 355–357; *Angew. Chem. Int. Ed. Engl.* **1971**, *10*, 330–331; b) K. C. Nicolaou, F. P. J. T. Rutjes, E. A. Theodorakis, J. Tiebes, M. Sato, E. Untersteller, *J. Am. Chem. Soc.* **1995**, *117*, 1173–1174.
- [20] a) B. O. Lindgren, T. Nilsson, *Acta Chem. Scand.* **1973**, *27*, 888–890; b) G. A. Kraus, M. J. Taschner, *J. Org. Chem.* **1980**, *45*, 1175–1176.
- [21] We thank Astellas Pharma for providing spectroscopic data of (+)-okilactomycin for comparison.
- [22] With regard to this synthesis of the unnatural enantiomer of **1a**, (–)-okilactomycin, since our intermediates up to and including **25** possessed (+) rotation values, we anticipated that the route would produce the natural (+) enantiomer. Comparing our structures to the previous synthesis by Smith (Ref. [2f]) was complicated since the absolute structure of (+)-**1a** in Figure 1 of Ref. [2f] should be the enantiomer. Importantly, all other structures and assignments in Ref. [2f] are correct (Prof. Amos Smith, private communication). Interestingly, both natural chrolactomycin and the recently isolated congeners of okilactomycin (okilactomycin A–D) have negative optical rotation values. Efforts are underway in our laboratory to produce the natural enantiomer along with structure analogues for our biological investigations.